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3 ORIGINAL PAPER

4 **Effect of dietary alternative lipid sources on haematological**
5 **parameters and serum constituents of *Heterobranchus***
6 ***longifilis* fingerlings**

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13 **Abstract** The worldwide increase in aquaculture
14 production and the decrease of wild fish stocks has
15 made the replacement of fish oil (FO) in aquafeed
16 industry a priority. Therefore, the use of terrestrial
17 animal fats and vegetable oils, which has lower cost
18 and larger supplies, may be good as substitute for FO.
19 This study investigate the effects of total replacement
20 of FO by two terrestrial animal fats (pork lard and
21 poultry fat) and three vegetable oils (palm kernel oil,
22 sheabutter oil and sunflower oil) on haematological
23 and serum biochemical profile of *Heterobranchus*
24 *longifilis* over 70 days. FO-diet was used as the
25 control. The haematological parameters were signifi-
26 cantly affected by dietary lipid sources. Serum total
27 protein was not influenced by the dietary lipids.
28 However, serum cholesterol was significantly higher
29 in fish fed diet containing sunflower oil. Glucose
30 and activities of liver enzymes in blood serum were

significantly reduced in fish fed alternative lipids 31
when compared with the control. These results indicate 32
that FO can be replaced completely with alternative 33
lipids without any serious negative health impacts. 34

Keywords Fish oil · Alternative lipids · Haematology · 35
Heterobranchus longifilis · Serum constituents 36

Introduction 37

Marine fish oils have traditionally been used in diets 38
for cultured fish to provide fish with energy and 39
essential fatty acids. However, these oils are in great 40
demand world-wide and as a result of their limited 41
supply, they are becoming increasingly more costly. 42
To maintain and enhance the economic viability of 43
aquaculture, it has become necessary to find suitable, 44
less expensive alternate plant and/or animal lipid 45
sources that would satisfy the nutritional requirement 46
of fish for growth and health. 47

Catfish can utilize saturated fats, and several 48
studies have focused on catfish growth rates as a 49
function of dietary lipid (Lim et al. 2001; Shirai et al. 50
2001; Ng et al. 2003). In these studies, no difference 51
was observed between fish fed fish oil or vegetable 52
oil or animal fats or their combinations. Differences in 53
haematological parameters, immune response and 54
disease resistance as a function of dietary lipid source 55
have also been reported for catfish (Fracalossi and 56
Lovell 1994; Klinger et al. 1996; Ochang et al. 2007), 57

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58 Atlantic salmon (Balfry et al. 2006) and large mouth
 59 bass (Subhadra et al. 2006). However, the influence of
 60 alternate dietary lipid sources on *Heterobranchus*
 61 *longifilis* haematological parameters and serum con-
 62 stituents are very scanty. Blood is a good indicator to
 63 determine the health of an organism. It also acts as
 64 pathological reflector of the status of exposed animal
 65 to toxicants (Joshi et al. 2002).

66 This study was initiated to determine if dietary
 67 alternate lipid sources, previously shown to affect
 68 haematological parameters of channel catfish, *Clarias*
 69 *gariiepinus*, Atlantic salmon and large mouth bass, had
 70 any effect on selected haematological parameters and
 71 serum constituents of *H. longifilis*. Total red blood cell
 72 (RBC) and white blood cell (WBC) counts were used
 73 as indicators of hemopoiesis. Measures of mean cell
 74 volume (MCV) allowed us to evaluate the physiologi-
 75 cal state of RBC. The haematological response of *H.*
 76 *longifilis* to dietary alternate lipid sources was
 77 evaluated by analyzing these characteristics.

78 **Materials and methods**

79 **Experimental diets**

80 The efficiency of five alternative lipid sources: two
 81 terrestrial animal fats (pork lard, PL and poultry fat,
 82 CF) and three vegetable oils (palm kernel oil, PKO;

sheabutter oil, SB and sunflower oil, SFO) were 83
 evaluated. Six isonitrogenous (450 gkg⁻¹) and iso- 84
 energetic (18 MJkg⁻¹) experimental diets were for- 85
 mulated (Table 1), containing 6% of added lipid 86
 sources. Fish oil (FO) was used in the control diet, 87
 which was completely replaced with the alternative 88
 lipid sources in the other five diets. The diets were 89
 made into pellet with meat mincer through 2 mm die, 90
 sundried, packed in polythene bags, sealed and stored 91
 at -20°C until used. 92

Animal and husbandry 93

Heterobranchus longifilis fingerlings were obtained 94
 from the hatchery of National Institute for Freshwater 95
 Fisheries Research (NIFFR), New-Bussa, Nigeria, 96
 and maintained in the laboratory for two weeks prior 97
 to the experiment. The fish were maintained in 98
 circular plastic tanks in a flow-through system during 99
 the acclimation and experimental period. The fish 100
 were reared under a natural photoperiod of 12D:12L. 101

During acclimation, fish were fed to apparent 102
 satiation twice a day using a commercial catfish diet 103
 (45 % CP, 12.5 % crude lipid and supply by NIFFR). 104
 Three tanks were randomly assigned to each diet 105
 group in 10 weeks experiment. At the start of the 106
 experiment, 20 fishes were batch weighed and 107
 stocked into each tank. During the feeding period, 108
 fish were fed the experimental diet to apparent 109

Table 1 Composition of the experimental diets (g/kg)

Ingredients	Diets						
	¹ FO	² SB	³ PKO	⁴ SFO	⁵ PL	⁶ CF	
Fish meal (Danish)	398	398	398	398	398	398	t1.4
Soybean meal	313	313	313	313	313	313	t1.5
Corn flour (Maize)	177	177	177	177	177	177	t1.6
Cassava starch	20	20	20	20	20	20	t1.7
Methionine	10	10	10	10	10	10	t1.8
⁷ Vit./Min. Premix	20	20	20	20	20	20	t1.9
Salt (NaCl)	1.5	1.5	1.5	1.5	1.5	1.5	t1.10
Vitamin C	0.5	0.5	0.5	0.5	0.5	0.5	t1.11
Fish oil	60						t1.12
(²⁻⁶)Alternative lipids		60	60	60	60	60	t1.13

¹ Fish oil (cod liver oil), ² Sheabutter oil, ³ Palmkernel oil, ⁴ Sunflower oil, ⁵ Pork lard, ⁶ Poultry fat, ⁷ Vitamin/mineral premix supplied t1.14
 the following (per kg of diet): calcium, 4500 mg; phosphorus, 4200 mg; potassium, 1700 mg; magnesium, 400 mg; iron, 30 mg; zinc,
 30 mg; manganese, 20 mg; copper, 5 mg; iodine, 1 mg; selenium, 0.25 mg; vitamin A, 5000 IU; vitamin D, 2000 IU; DL- α -
 tocopherol acetate, 100 mg; menadione, 15 mg; thiamine hydrochloride, 5 mg; riboflavin, 10 mg; pyridoxine hydrochloride, 10 mg.
 Panthothenic acid, 35 mg; nicotinic acid, 50 mg; biotin, 0.5 mg; folic cid, 2 mg; ascorbic acid, 200 mg; inositol, 250 mg; choline,
 400 mg; vitamin B₁₂, 0.1 mg and ethoxyquin, 60 mg.

110	satiation twice a day at 09:00 and 16:00 respectively.	analyzed by using commercial clinical investigation kits (Wako, Osaka, Japan).	153 154
111	One hour, later uneaten feed was siphoned, dried to		
112	constant weight at 70°C and reweighed.		
113	Sample collection	Statistical analysis	155
114	At the end of the trial, fish were tranquilized with	The data were subjected to analysis of variance (ANOVA) and if significant (P<0.05) differences were found, Duncan's multiple range test (Duncan 1955) was used to rank the group using SPSS version 10.0 (SPSS 1997). The data presented as mean ± S.E. M. of three replicate groups.	156 157 158 159 160 161
115	150 mg l ⁻¹ solution of methane sulphonate (MS222)		
116	(Wagner et al. 1997) for blood collection. Blood		
117	samples were obtained from the caudal vein of five		
118	fish from each tank. 1 ml blood sample was collected		
119	into bottles containing 0.05 ml EDTA as anticoagu-		
120	lant. Blood samples for serum analysis were collected		
121	into bottles without any anticoagulant. Serum was		
122	separated by centrifugation at 3500 x g for 5 minutes,		
123	and kept frozen at-20°C for the determination of		
124	protein, cholesterol, glucose, aspartate aminotransferase		
125	(AST) and alanine aminotransferase (ALT) activities.		
126	Haematological and blood serum biochemical profile	Results	162
127	Immediately after sampling, blood smears were	Haematological profile	163
128	prepared, red blood and white blood cell counts were	The mean value for the haematological parameters of <i>Heterobranchus longifilis</i> fingerlings studied are shown in Table 2. Fish fed diets containing SFO and CF had increased PCV, Hb, RBC and WBC contents than those on the other diet groups. Red blood cell and haemoglobin content for fish fed the control (FO), SB and PKO diets were similar.	164 165 166 167 168 169 170
129	carried out using standard haematological techniques	The blood indices calculated from the mean values of blood parameters shows that MCH and MCHC were significantly reduced in <i>H. longifilis</i> fed on diets containing FO, SB, PKO, SFO and PL. While MCV was significantly increased in fish fed diets containing SFO and PL. Percentage of neutrophils and lymphocytes in <i>H. longifilis</i> fed FO was significantly different from the other groups. The blood platelets were significantly different among fish fed different lipid sources.	171 172 173 174 175 176 177 178 179 180
130	(Dacie and Lewis 2001). 50 µl haematocrit tube was		
131	filled with blood samples, after centrifugation (3500 x		
132	g for 10 min) of each blood sample, packed cell		
133	volume (PCV) was determined by the Wintrobe and		
134	Westergreen method as described by Blaxhall and		
135	Daisley (1973). Haemoglobin (Hb) concentration esti-		
136	mates were determined as described by Wedemeyer		
137	and Yasutake (1977). Measurement of red blood cell		
138	count (RBC), haemoglobin concentration (Hb) and		
139	PCV enabled the mean cell volume (MCV), mean		
140	cellular haemoglobin content (MCH) and mean cell		
141	haemoglobin concentration (MCHC) to be calculated		
142	according to the following formulas (Dacie and Lewis		
143	2001):	Blood serum biochemical profile	181
	$MCV(fl) = PCV/RBC(10^6\mu l^{-1})$	Total protein, total cholesterol and glucose concentrations, and AST and ALT activities in serum of <i>H. longifilis</i> at the end of feeding trial are shown in Table 3. Serum total protein of fish was not significantly affected by dietary lipid sources. Serum glucose and total cholesterol were significantly higher in <i>H. longifilis</i> fed diets containing SFO and PL.	182 183 184 185 186 187 188
	$MCH(pg) = [Hb(gdl^{-1}) \times 10]/RBC(10^6\mu l^{-1})$	Serum AST activity increased significantly in <i>H. longifilis</i> fed alternative lipid diets compared to the control, except for those fed PKO that had significantly lower value. The highest value was obtained in group fed SB diets. A significantly reduction in ALT	189 190 191 192 193
148	and		
	$MCHC(gl^{-1}) = [Hb(gdl^{-1}) \times 10]/PCV$		
151	The concentrations of total protein, total cholesterol,		
152	glucose, AST and ALT activities in plasma were		

Table 2 Haematological profile of *Heterobranchus longifilis* fingerlings fed diets containing alternative lipid sources for 10 weeks

Parameters	FO	SB	PKO	SFO	PL	CF
PCV (%)	28.00±0.58 ^a	31.00±0.55 ^b	28.00±0.56 ^a	35.00±0.58 ^c	28.00±0.59 ^a	31.00±0.58 ^b
Hb (gdL ⁻¹)	10.80±0.06 ^b	10.20±0.06 ^b	10.10±0.06 ^b	11.20±0.12 ^c	9.00±0.23 ^a	12.50±0.01 ^c
RBC (X 10 ⁶ µL ⁻¹)	2.42±0.01 ^b	2.49±0.02 ^b	2.37±0.01 ^b	2.63±0.02 ^c	2.03±0.02 ^a	2.62±0.01 ^c
MCV (fl)	117.33±0.88 ^a	123.00±0.58 ^b	118.00±0.58 ^a	132.00±0.58 ^c	135.00±1.73 ^c	117.00±0.58 ^a
MCH (pg)	41.00±0.58 ^a	42.00±1.51 ^a	43.00±1.73 ^a	43.00±0.58 ^a	44.00±2.31 ^{ab}	48.00±0.01 ^b
MCHC (g ⁻¹)	37.00±1.51 ^{bc}	35.00±1.73 ^{ab}	36.00±2.31 ^{ab}	32.00±0.58 ^a	32.00±0.58 ^a	41.00±0.58 ^c
WBC (X 10 ⁹ µL ⁻¹)	220.00±1.15 ^a	244.00±1.15 ^b	222.00±1.15 ^a	250.00±0.58 ^c	220.00±1.15 ^a	249.00±0.58 ^c
Neutrophils (%)	5.00±0.58 ^b	2.00±0.07 ^a	2.00±0.59 ^a	2.00±0.58 ^a	3.00±0.58 ^a	3.00±0.56 ^a
Lymphocytes (%)	95.00±0.58 ^a	98.00±0.57 ^b	98.00±0.59 ^b	98.00±0.58 ^b	97.00±0.58 ^b	97.00±0.56 ^b
Platelets	210.00±1.15 ^a	312.00±1.15 ^d	263.00±1.73 ^b	329.00±1.15 ^c	250.00±1.15 ^b	285.00±0.58 ^c

Data are mean ± S.E.M. (n=3): means with different superscripts are significantly different (P<0.05).

194 activity was recorded in group fed the control diet.
 195 The CF diet yielded a significantly higher (P<0.05)
 196 ALT than in other dietary groups.

197 **Discussions**

198 Fish haematology is gaining increasing importance in
 199 fish culture because of its importance in monitoring
 200 the health status of fish (Hrubec et al. 2000). Results
 201 of the haematological parameters of *H. longifilis* in
 202 this study showed that there were significant differ-
 203 ences (P<0.05) among different dietary groups. It has
 204 been shown that variables such as age, sex, dietary
 205 state and stress alter blood values (Barnhart 1969;
 206 McCarthy et al. 1973). Haemoglobin concentration
 207 and PCV of fish blood has also been shown to
 208 decrease after the capture and transportation (Hattingh
 209 and Van Pletzen 1974). The fish used in this study
 210 were kept under laboratory condition during the period
 211 of feeding with test diets and were tranquilized with
 212 MS222 before handling. The effect of stress resulting

from handling must have been minimal. Consequently,
 only the nutritional states of the fish have significantly
 affected the haematological parameters. However, the
 PCV values obtained in this study higher than the
 values obtained for *Clarias gariepinus* (Ochang et al.
 2007) and bagrid catfish (Etim et al. 1999).

Similarly, the Hb content of fish in this study
 exhibited higher values than those obtained by
 Subhadra et al. (2006) for large mouth bass with diets
 containing canola oil, chicken oil and menhaden fish
 oil, which ranged between 3.7–3.9 g dL⁻¹. This shows
 that the oxygen carrying abilities of the blood of *H.*
longifilis higher, and could be attributed to species
 differences and their ability to utilize n-6 fatty acids
 present in vegetable oils.

The lower values of RBC and WBC in fish fed FO
 show clearly that the African catfish cannot effective-
 ly utilize n-3 PUFA without dilution with n-6 PUFA.
 The n-3:n-6 PUFA balance seems critical in the diet
 of African catfish (Ochang et al. 2007). It is likely
 that n-3 PUFA in the cod liver oil may have significantly
 affected the white blood cells and other blood param-

Table 3 Blood serum biochemical profile of *Heterobranchus longifilis* fingerlings fed diets containing alternative lipid sources for 10 weeks

Diets	Serum total protein (g/100 ml)	Serum total cholesterol (mg/100 ml)	Serum glucose (mg/100 ml)	Serum AST (IU/l)	Serum ALT (IU/l)
FO	2.86±0.08	35.47±0.33 ^c	26.21±0.31 ^a	16.11±0.06 ^b	10.11±.05 ^a
SB	2.72±0.11	41.97±0.55 ^c	26.33±0.22 ^a	39.08±0.05 ^c	12.12±0.06 ^b
PKO	2.75±0.08	30.80±0.46 ^a	26.79±0.22 ^b	10.05±0.04 ^a	17.10±0.05 ^d
SFO	2.93±0.09	46.73±0.50 ^f	27.18±0.15 ^c	20.01±0.01 ^c	16.04±0.02 ^c
PL	2.90±0.07	37.37±0.27 ^d	27.29±0.15 ^c	22.03±0.02 ^d	16.08±0.05 ^c
CF	2.81±0.06	31.83±0.43 ^b	26.20±0.16 ^a	13.07±0.04 ^{ab}	18.08±0.04 ^c

Data are mean ± S.E.M. (n=3): means with different superscripts are significantly different (P<0.05).

eters, thereby compromising immune system. This may explain lower percent lymphocytes recorded for fish fed FO diet. Lymphocytes are responsible for immune responses and neutrophil is a form of granulocyte that kills and digests microorganisms it has engulfed by phagocytosis. Lall (2000) revealed that fatty acid composition of diet influences production of eicosanoids precursors and immune response in laboratory animals. A diet high in n-6 PUFAs produces relatively high levels of pro-inflammatory 2-series prostaglandins (PGs) and 4-series leucotrienes (LTs) and lipoxins (LXs) derived from arachidonic acid (AA) whereas diets high in n-3 PUFAs produce the anti-inflammatory 3-series PGs and 5-series LTs and LXs derived from eicosapentaenoic acid (EPA). Generally, diets containing n-6 PUFAs enhance the immune response due to the high levels of pro-inflammatory AA-derived eicosanoids, and diet containing n-3 (FO) may be immunosuppressive due to the high levels of EPA-derived anti-inflammatory eicosanoids.

Blood indices (MCV, MCH and MCHC) are particularly important for the diagnosis of anaemia in most animals (Coles 1986). This study showed a significant decrease of MCV and MCH in fish fed FO. So, it is assumed that the decrease or increase of blood indices suggest a possible hemoconcentration, a situation which coincided with the main changes observed in *Prochilodus scrofa* exposed to copper (Mazon et al. 2002).

The reduction in platelets reported in this study for fish fed FO compared with that in fish fed with the alternative lipids may indicate that oil rich in n-3 highly unsaturated fatty acids (HUFAs) (such as FO) reduces the blood clotting tendency and minimises inflammatory responses as suggested by Pepping (1999) and Vauy and Valenzuele (2002).

Blood serum protein is fairly labile biochemical system, reflecting the condition of the organism and the changes happening to it under the influence of internal and external factors. Booke (1964) showed that sex, spawning, food, osmotic pressure, temperature, light, age, hibernation hormones, oxygen depletion and season are factors that demand total serum protein complement in fish. In this study, no significant hyperproteinemia was observed in all fish groups fed various alternative lipids. This shows that the various lipid sources do not lead to osmoregulatory dysfunction, hemodilution or damage of tissues surrounding blood vessels of *H. longifilis*. Hille (1982) attributed high

serum protein in rainbow trout to be an indicative of osmoregulatory dysfunction, damage of tissues surrounding blood vessels and hemodilution.

Larsson and Fange (1977) reported that the normal plasma cholesterol values ranging from 86 to 921 mg/100 ml in most marine fish were two to six times higher than that of mammalian plasma. Similarly, low plasma cholesterol concentration has been reported in Japanese flounder fed diet containing lauric acid as sole lipid source (Kim et al. 2002) and starry flounder (Lee et al. 2003).

In the present study, serum glucose concentration reduced significantly in fish fed diets containing cod liver oil, SB and CF. on the other hand the elevated ($P < 0.05$) serum glucose in fish fed PKO, SFO and PL, indicate that these lipid sources affects glucose dynamics in *H. longifilis* in order to obtain more energy to withstand and overcome existing stress condition. Plasma or serum glucose levels are often used as an indicator of non-specific stress (Hunn and Greer 1991). Increased blood serum glucose levels might have been due to a glucose shift from tissue to blood or to an impairment of glucose mobilization.

Transamination represents one of the main pathways for synthesis and deamination of amino acids, thereby allowing interplay between carbohydrate and protein metabolism during the fluctuating energy demands of the organism in various adaptive situations. It is also considered to be important in assessing the state of the liver and some other organs (Verma et al. 1981). Therefore attention has been focused on the changes in AST, ALT and alkaline phosphate (ALP) activities, which promotes gluconeogenesis from amino acid, as well as on the changes in aminotransferase activities in the liver (Hilmy et al. 1981; Rashatur and Ilyas, 1983). Furthermore, AST and ALT might be altered by a variety of chemical, biological and physiological factors or by a disturbance in the Krebs cycle. Decreased activity of the Krebs cycle cause a decrease in its intermediates, thereby, ALT and AST compensate by providing α -ketoglutarate (Salah El-Deen and Rogeps 1993). Results of this study showed that serum AST and ALT activities increased significantly in the fish group fed on alternative lipid sources-based diets. The elevated serum AST in *H. longifilis* fed SB and PL is probably due to release of transaminase from cytoplasm due to hepatic cellular damage. It has been reported that higher GOT activities were found in rock fish (Lee 2001), Japanese flounder (Kim et al.

333 2002) and starry flounder (Lee et al. 2003) fed the n-3
 334 HUFA-deficient diets. However, AST and ALT values
 335 found were within the range consistent with good fish
 336 health (Sandnes et al. 1988).
 337 Under the experimental conditions described in this
 338 study, any of the diet treatment had serious negative
 339 health impacts. It is feasible to completely replace FO
 340 with alternative lipids in the diets of *H. longifilis*
 341 without any negative affects on haematological and
 342 serum biochemical profile of the fish.

344 **References**

345 Balfray, S.K.; Oakes J.; Rowshandeli, M.; Deacon, G.; Skura,
 346 B.J.; Higgs D.A., 2006: Efficacy of equal blend of canola
 347 oil and poultry fat as an alternate dietary lipid source for
 348 Atlantic salmon (*Salmo salar* L.) in sea water. II: effects
 349 on haematology and immunocompetence. *Aquaculture*
 350 *Res.* 37: 192–199. doi:10.1111/j.1365-2109.2005.01421.x
 351 Barnhart, R.A., 1969: Effects of certain variables on haemato-
 352 logical characteristics of rainbow trout, *Salmo gairdneri*
 353 (Richardson). *Trans. Am. Fish Soc.* 98: 411–418.
 354 doi:10.1577/1548-8659(1969)98[411:EOCVOH]2.0.CO;2
 355 Blaxhall, P.C.; Daisley, K.W., 1973: Routine haematological
 356 methods for use with fish blood. *J. fish Biol.* 5: 771–781.
 357 Booke, H.E., 1964: A review of variations found in fish serum
 358 proteins. *New York Fish Game J.* 11: 47–57.
 359 Coles, E.H., 1986: Veterinary clinical pathology. Philadelphia:
 360 Saunders, 615p.
 361 Dacie, J.V.; Lewis, S.M., 2001: Practical Haematology 9th ed.
 362 Churchill Livingstone, London, 633 pp.
 363 Duncan, D.B., 1955: Multiple range and multiple (F) test.
 364 *Biometrics*, 11:1–42. doi:10.2307/3001478
 365 Etim, L.; Ekanem, S.B.; Utin, A., 1999: Haematological profile
 366 of two species of catfish. *Chrysichthys nigrodigitatus*
 367 (Lacepede) and *Chrysichthys furcatus* (Gunter) from the
 368 Great Kwa River, Nigeria. *Global J. Pure and Appl. Sci.*,
 369 5:1–4.
 370 Fracalossi, D.M.; Lovell, R.T., 1994: Dietary lipid sources influence
 371 responses of channel catfish (*Ictalurus punctatus*) to challenge
 372 test with the pathogen *Edwardsiella ictaluri*. *Aquaculture*
 373 119: 287– 298. doi:10.1016/0044-8486(94)90183-X
 374 Hattingh J.; Van Pletzen A.J., 1974: The influence of capture
 375 and transportation on some blood parameter of freshwater
 376 fish. *Comp. Biochem physiol.* 49a: 607–609.
 377 Hille, S., 1982: A literature review of the blood chemistry of
 378 rainbow trout, *Salmo gairdneri* Rich. *J. Fish. Biol.* 20:
 379 535–569. doi:10.1111/j.1095-8649.1982.tb03954.x
 380 Hilmy, A.M.; Shabana, M.B.; Said, M.M., 1981: The role of
 381 serum transaminase (SGOT and SGPT) and alkaline
 382 phosphates in relation to organic phosphorus with respect
 383 to mercury poisoning in *Alphanius dispar* Rupp (Teleost)
 384 of the red sea. *Comp. Biochem. Physiol.* 68: 69–74. doi:
 385 10.1016/0306-4492(81)90039-3
 386 Hrubec, T.C.; Cardinale, J.L.; Smith, S.A., 2000: Hematology
 387 and plasma chemistry reference intervals for cultured

Tilapia (Oreochromis hybrid). *Verter. Clin. Path.* 29: 7– 388
 12. doi:10.1111/j.1939-165X.2000.tb00389.x 389
 Hunn, J.B.; Greer, I.E., 1991: Influence of sampling on the 390
 blood chemistry of Atlantic salmon. *Prog. Fish Cult.* 53: 391
 184–187. doi:10.1577/1548-8640(1991)053<0184: 392
 IOSOTB>2.3.CO;2 393
 Joshi, P.K.; Bose, M.; Haris, D., 2002: Changes in certain 394
 haematological parameters in a silurid catfish *Clarias* 395
batrachus (Linn) exposed to cadmium chloride. *Pollution* 396
Resources 21 2: 129–131. 397
 Kim, K.-D.; Lee, S.-M.; Park, H.G.; Bai, S.; Lee, Y.-H., 2002: 398
 Essentiality of dietary n-3 highly unsaturated fatty acids in 399
 juvenile Japanese flounder (*Paralichthys olivaceus*). *J.* 400
World Aquac. Soc. 33, 432– 440. doi:10.1111/j.1749-7345. 401
 2002.tb00022.x 402
 Klinger, R.C.; Blazer, V.S.; Echevarria, C., 1996: Effects of 403
 dietary lipid on the haematology of channel catfish, *Ictalurus* 404
punctatus. *Aquacult.* 147: 335–233. doi:10.1016/S0044- 405
 8486(96)01410-X 406
 Lall, S.P., 2000: Nutrition and health of fish. In Cruz-Suárez, 407
 zL.E., Ricque-Marie, D., Tapia-Salazar, M., Olivera-Nova, 408
 M.A., Civera-Cerecedo, R. (Eds.). avances en Nutrición 409
 Acuicola V. Memorias del V Simposium Internacional de 410
 Nutrición Acuicola. 19–22 Noviembre, 2000. Mérida, 411
 Yucatán, Mexico. 412
 Larsson, A.; Fange, R., 1977: Cholesterol and free fatty acids 413
 (FFA) in the blood of marine fish. *Comp. Biochem. Physiol.* 414
 57B: 191–196. 415
 Lee, S.-M., 2001: Review of the lipid and essential fatty acid 416
 requirements of rockfish (*Sebastes schlegeli*). *Aquac. Res.* 417
 32 Suppl. 1, 8 –17. 418
 Lee, S.-M.; Lee, J. H.; Kim, K.-D., 2003: Effect of dietary 419
 essential fatty acids on growth, body composition and 420
 blood chemistry of juvenile starry flounder (*Platichthys* 421
stellatus). *Aquaculture* 225: 269–281 doi:10.1016/S0044- 422
 8486(03)00295-3 423
 Lim, P.-K.; Boey, P.-L.; Ng, W.-K., 2001: Dietary palm oil level 424
 affects growth performance, protein retention and tissue 425
 vitamin E concentration of African catfish, *Clarias* 426
gariepinus *Aquaculture* 202:101–112 doi:10.1016/S0044- 427
 8486(01)00563-4 428
 Mazon, A.F.; Monteiro, E.A.S.; Pinheiro, G.H.D.; Fernandes, 429
 M.N., 2002: Hematological and physiological changes 430
 induced by short-term exposure to copper in the freshwa- 431
 ter fish, *Prochilodus scrofa*. *Braz. J. Biol.* 62: 621–631 432
 doi:10.1590/S1519-69842002000400010 433
 McCarthy, D.H.; Stevensom, J.P.; Roberts, M.S., 1973: Some 434
 blood parameters of the rainbow trout (*Salmo gairdneri* 435
Richardson). I. The kamloops variety. *J. Fish Biol* 5: 1–8. 436
 doi:10.1111/j.1095-8649.1973.tb04425.x 437
 Ng, W.-K.; Lim, P.-K.; Boey, P.-L., 2003: Dietary lipid and palm 438
 oil source affects growth, fatty acid composition and muscle 439
 a-tocopherol concentration of African catfish, *Clarias* 440
gariepinus. *Aquaculture* 215: 229–243 doi:10.1016/S0044- 441
 8486(02)00067-4 442
 Ochang, S. N.; Oyedapo, A.; Fagbenro; Olabode, T. A., 2007: 443
 Growth Performance, Body Composition, Haematology 444
 and Product Quality of the African Catfish (*Clarias* 445
gariepinus) Fed Diets with Palm Oil. *Pak. J. Nut.* 6 5: 446
 452–459, 2007 447

- 448 Pepping, J., 1999: Omega-3 essential fatty acids. *American*
449 *Journal of Health System Pharmacy* 56: 719–722. 469
- 450 Rashatuar, S.S.; Ilyas, R., 1983: Effect of chronic herbicide
451 intoxication on *in vivo* activities of certain enzymes in the
452 liver of freshwater fish *Nemachelius denisonii* (day). *Toxicol.*
453 *Lett.* 16: 249–252. doi:10.1016/0378-4274(83)90184-4 470
- 454 Subhadra, B.; Lochmann, R.; Rawles, S.; Chen, R., 2006:
455 Effect of dietary lipid source on the growth, tissue
456 composition and hematological parameters of largemouth
457 bass (*Micropterus salmoides*). *Aquaculture* 255: 210–222.
458 doi:10.1016/j.aquaculture.2005.11.043 471
- 459 Salah El-Deen, M.; Rogeps, W. A., 1993: Changes in total
460 protein and transaminase activities of grass carp exposed to
461 diquat. *J. Aquatic Animal Health*, 5: 280–286. doi:10.1577/
462 1548-8667(1993)005<0280:CITPAT>2.3.CO;2 472
- 463 Sandnes, K.; Lie, Ø.; Waagbø, R., 1988: Normal ranges of
464 some blood chemistry parameters in adult farmed Atlantic
465 salmon, *Salmo salar*. *J. Fish Biol.*, 32: 129–136. doi:
466 10.1111/j.1095-8649.1988.tb05341.x 473
- 467 Shirai, N.; Suzuki, H.; Toukairin, S.; Wada, S., 2001: Effect of
468 Japanese catfish *Silurus asotus* lipid intake on the lipid
469 components of plasma and liver in adult mice. *Fisheries*
470 *Science.* 67: 321–327. doi:10.1046/j.1444-2906.2001.00230.x 471
- SPSS, 1997: SPSS Base 7.5 for Window. SPSS, 444 N. 472
- Michigan Avenue, Chicago, IL, USA. 473
- Vaay, D.R.; Valenzuele, A., 2002: Marine oils. The health
474 benefits of n-3 fatty acids. *Nutrition Reviews* 54: 102–108. 475
- Verma, S.R.; Rani, S.; Delela, R.C., 1981: Isolated and
476 combined effects of pesticides on serum transaminases in
477 *Mystus vittatus* (African catfish). *Toxicol. Lett.* 8: 67–71.
478 doi:10.1016/0378-4274(81)90140-5 479
- Wagner, E. J.; Jessen, T.; Arndt, R.; Routledge, M.D.; 480
- Brddwisch, Q., 1997: Effects of rearing density upon
481 cutthroat trout hematology, hatchery performance, fin
482 erosion and general health and condition. *The Progressive*
483 *Fish-Culturist* 59: 173–187. doi:10.1577/1548-8640(1997)
484 059<0173:EORDUC>2.3.CO;2 485
- Wedemeyer, G.A.; Yasutake, W. T., 1977: Clinical methods for
486 the assessment of the effects of environmental stress on
487 fish health. Technical papers of the US Fish and Wildlife
488 Service No 89. Washington, DC. US Dept. of the Interior,
489 Fish and Wildlife Service 18p. 490

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